

**Changes in Some Hematological Parameters of Female Patients on Directly Observed Treatment Short-Course (DOTS) in a Tuberculosis Hospital in Bayelsa State, Nigeria**

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**ABSTRACT**

**Introduction:** Tuberculosis disease remains a health challenge globally. Anti-TB drugs are not completely free from side effects. **Objective;** This study was aimed at determining the effect/s of first and second line Directly Observed Treatment Short-course (DOTS) therapy on the selected hematological parameters of female TB infected patients. **Materials and methods.** The longitudinal study involved 198 female subjects attending a TB hospital in Bayelsa State, Nigeria. AFB test was done using Zeihl-Neelson technique, ESR by the westerngreen method and hematological parameters analyzed with the Abacus 380 Hematology Autoanalyzer. ANOVA was used to compare the mean values and SPSS version 20.00 was employed to analyze the data. **Results:** Results from the study indicate that there was significant difference in most parameters when control values were compared with that of the newly infected TB subjects. They include; WBC (p=0.0001), RBC (p=0.0001), HB (p=0.0001), PCV (p=0.0001) and ESR (p=0.0001). On the post-hoc involving control/second line treatment subjects, five items yielded insignificant values; WBC, RBC, PCV, HB and MCH. **Conclusion:** While our study showed that TB disease significantly affected all the parameters, there was a real improvement in the treatment stage as all parameters were returning back to normal values. This suggests that as time progresses absolute normalization can occur. It is inferred from this that the DOTS chemotherapy has no negative effect on the hematopoietic process. We therefore suggest further studies that will look into the plasma components of patients undergoing DOTS chemotherapy.

**Key words;** Anti-TB Therapy, DOTS, Hematological Parameters,

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## INTRODUCTION

Tuberculosis also known as TB, is a major health issue and a principal infectious killer disease. It is caused mainly in man by the parasite *Mycobacterium tuberculosis*. The bacillus enters the body by breathing of droplets or dust particles containing it. It occurs as pulmonary tuberculosis or non-pulmonary tuberculosis(1). It is known globally that 700 000 women die from tuberculosis every year; this infectious ailment kills more females than do all causes of maternal mortality combined.

In Africa (especially sub-Saharan Africa), cases of tuberculosis have grown dramatically, overpowering control programs (2). This negative effect could be due to the fact that tuberculosis infection control interventions are not routinely implemented as against what happens in high income countries with low prevalence of tuberculosis where infection control policies are routinely observed(3). However, the launch of the Directly Observed Treatment Short-course (DOTS) in 1995 by the World Health Organization (WHO) has shown to be an actual intervention that will lead to reduced tuberculosis transmission and lessening number of tuberculosis cases(4)(5). Reports have shown that it is among the most economical global health interventions available today(6)(7). DOTS was first implemented in India in the year 1958. It is made up of five components which include (1) government obligation to control activities. (2) Case detection by sputum smear microscopy. (3) Standardized treatment regimens lasting at 6 – 8 months directly observed for 2 months. (4) Regular and continual provision of anti-TB drugs and (5) standardized recording and reporting system. The major objective behind this strategy is to form a close relationship between patients and health workers or

volunteers in order to help them successfully complete treatment, and to avert drug resistance (8). DOTS extension and improvement remains one of the vital tactics adopted by the National TB and Leprosy Control Program to the realization of a general access to TB services in Nigeria (9). Several research works have revealed significant alterations in hematological parameters amongst subjects with new cases of TB compared with healthy subjects (10, 11, 12, 13, 14, 15, 16). However, there are very few researches that describes the **hematological** parameters of infected females throughout the 6 or 9-month period of administration of the DOTS treatment with only some centering only on the first line course of therapy (17)(18). This study was aimed at determining the effect of first line and second line DOTS therapy on the **hematological** parameters of Pulmonary TB infected female patients.

## MATERIALS AND METHODS

### Subjects and Setting

A Longitudinal study design was adopted in this study; involving 198 female subjects. Subjects were congregated into four (4) groups: (1) Apparently healthy control subjects, (2) Newly diagnosed TB subjects, (3) First line (Intensive Phase) DOTS treatment (Isoniazid, Rifampin, Pyrazinamide and Ethambutol) subjects who are at the end of their second (2<sup>nd</sup>) month treatment and (4) Second line (Continuation Phase) DOTS treatment (Rifampisin and Isoniazide), subjects who are at the end of their six (6<sup>th</sup>) month treatment. Their ages ranged from 25 to 50 years. Smokers, alcohol consumer, persons with septic wounds, people with diabetes and other chronic illness, were excluded from the study. The study was carried out at the Tuberculosis and leprosy Hospital Igbogone, Yenagoa Local

Government Area of Bayelsa State Nigeria. Authorization for the study was gotten from the ministry of Health Bayelsa State Nigeria with ethical approval number BSMoHEC/2018/015 on 21<sup>st</sup> October 2018.

### **Blood Sample Collection**

Blood sample was collected as described by Ochei and Kolhatkar (1). Five (5) ml of venous blood was collected from each of the subjects and carefully put into Ethylene Diamine Tetra Acetic acid (EDTA) containers. After proper mixing, the samples were analyzed.

**Tuberculosis Test:** The Ziehl Neelsen method was used to detect Acid Fast Bacilli (AFB). Infection was confirmed by microscopic identification of tuberculosis bacilli in stained sputum smear (1).

### **Full Blood Count Analysis**

Full Blood Count Analysis was done with the Abacus 380 Hematology Auto analyzer. The items analyzed were White Blood Count (WBC), Red Blood Count (RBC), Packed Cell Volume (PCV) red cell indices of Mean Cell Volume (MCV), Mean Cell Hemoglobin (MCH) and Mean Cell Hemoglobin Concentration and hemoglobin estimation (HB). The operating principle of this analyzer is the impedance or coulter method (19). In this method, individual cell is suspended in a conductive diluent acts as an insulator. As the cells go through an aperture, they increase the resistance of the electrical path between the

submerged electrodes on either side of the aperture. Consequently, a measurable electronic pulse is generated. The number of pulses is proportional to the number of cell particles, and cell volume (20).

For the Hemoglobin measurement, the sample is lysed, diluted and measured by photometry. The lytic process releases hemoglobin. This hemoglobin is converted chemically, to methemoglobin which is measured by a photometer in the chamber (19).

### **Erythrocyte Sedimentation Rate (ESR)**

ESR was estimated by the westerngreen method (1).

### **Statistical analysis:**

Data collected from this research work was analyzed with **SPSS version 20.00** software. The inferential statistics Analysis of Variance (ANOVA) was employed to compare the parameters under study. A p value of less than 0.05 ( $p < 0.05$ ) was considered significant.

## **RESULTS**

The total number who partook in the study was 198 female subjects. They were clustered into four categories: Control group (58), New cases (38) First line treatment group (36) and Second Line treatment TB subjects (66) (Figure 1). Their age ranged from 25 to 50 years. For all groups. Scientific comparisons were made between each group.



**Figure 1. Distribution of subjects**

In Table 1., the mean values for all categories is compared using Analysis of variance (ANOVA). One (1) parameter had insignificant p values while the rest seven (7) had significant p values. The insignificant one is; MCH (f = 0.70 and p = 0.5590). The others with significant values include RBC (f

= 8.081 and p = 0.0001), ESR (f = 37.45 and p = 0.0001), MCV (f = 443.19 and p = 0.0001), PCV (f = 9.865 and p = 0.0001), HB (f = 8.509 and p = 0.0001), MCHC (f = 18.38 and p = 0.0001) and lastly WBC (f= 27.02 and p= 0.0001).

**Table 1a. Comparison of Mean ± SD of Hematological Parameters among Control, New cases, First line and Second line female subjects in TB treatment.**

Parameter		Control Subjects	New cases	First line	Second line	f, df	p-value
	N =	58	38	36	66		
<b>WBC(x10<sup>9</sup>/L)</b>	Mean±SD	6.16±1.38	8.67±2.44	6.57±1.36	5.69±1.56	27.02,3	0.0001*
<b>RBC(x10<sup>12</sup>/L)</b>	Mean±SD	4.90±0.72	4.28±0.31	4.56±0.71	4.60±0.57	8.081,3	0.0001*
<b>HB (g/dl)</b>	Mean±SD	12.06±1.67	10.36±1.77	11.14±1.85	11.44±1.40	8.509,3	0.0001*
<b>PCV (%)</b>	Mean±SD	36.75±5.41	31.06±5.10	33.72± 6.20	34.97±1.18	9.865,3	0.0001*
<b>ESR (mm/h)</b>	Mean±SD	6.27 ± 1.09	54.36±39.01	61.44±36.76	58.03±36.53	37.45,3	0.0001*
<b>MCV (fl)</b>	Mean±SD	74.90±5.80	72.84± 8.20	74.28± 5.32	78.42±5.52	443.19,3	0.0001*
<b>MCH (pg)</b>	Mean±SD	24.71±1.93	24.02± 2.81	24.39± 1.72	24.43±2.44	0.70,3	0.5590
<b>MCHC (g/dl)</b>	Mean±SD	329.24±6.95	330.53±7.87	328.39±6.96	321.03±8.18	18.38,3	0.0001*

**Key:** WBC: White Blood Count, RBC: Red Blood Count, HB: Hemoglobin concentration, PCV: Packed Cell Volume, ESR: Erythrocyte Sedimentation Rate, MCV: Mean Cell Volume, MCH: Mean Cell Hemoglobin, MCHC: Mean Cell Hemoglobin Concentration. \*Significant value

Table 1b shows the Post Hoc analysis of Control/New cases and Control / First line subjects. The Mean±SD of Control/New cases for WBC, RBC, HB, PCV and ESR are 6.16±1.38/8.67±2.44, 4.90±0.72/4.28±0.31, 12.06±1.67/10.36±1.77, 36.75±5.41/31.06±5.10 and 6.27±1.09/54.36±39.001 respectively. The post-hoc gave a P value of p= 0.0001 for all parameters.

The other parameters i.e. MCV, MCH and MCHC all had insignificant values. For MCV, the Mean±SD of Control/New cases was 74.90±5.80/72.84±8.20. The post-hoc gave a p value of p=0.4900. That of MCH and MCHC are 24.71±1.93/24.02±2.81 and 329.24±6.95/330.53±7.87 with a post- hoc of p= 0.5500 and p = 0.8820 respectively.

The second part of the table is the post hoc of Control/Firstline subjects. The parameters whose comparison of the control and firstline values was significant are ESR (p= 0.0001) and MCV (p = 0.0001). The rest parameters which include WBC, RBC, HB, PCV, MCH and MCHC all had their p values greater than 0.050(0.7190,0.0780, 0.0780, 0.550, 0.9350 and 0.9630 respectively).

Table 1c contains the post hoc analysis of Control/ second line and New cases / First line subjects. On the first part of the table, the control subjects had their WBC values higher than those on second line treatment (6.16±1.38/5.69±1.56). The post hoc analysis gave a p value of p= 0.4920 which is insignificant. Furthermore, it reveals an insignificant p values for RBC (p=0.0690), HB (p=0.2390), PCV (p= 0.2950) and MCH (p= 0.9010). The post hoc values for the other remaining three parameters were significant; ESR (p = 0.0001) MCV (p= 0.0240) and MCHC (p= 0.0001).

The second part of the table gives the post hoc p values for New cases / First line subjects. For WBC, the value for New case is 8.67±2.44 as compared to first line value of 6.57±1.36 which resulted to a significant p-value of p= 0.0001. Another significant p value was seen in MCV (72.84±8.20/74.28±5.32, p=0.0001). The table further reveals all other parameters as having insignificant p values. They include; RBC p = 2870, HB p= 0.2400, PCV p= 0.1770, ESR p= 0.8150, MCH0 p= 9210 and MCHC p= 0.6880.

**Table 1b. Post- Hoc analysis of Control/New cases and Control / First line subjects**

Parameter		Control Subjects	New cases	p-value	Control Subjects	First line	p-value
	N =	58	38		58	36	
<b>WBC(x10<sup>9</sup>/L)</b>	Mean±SD	6.16±1.38	8.67±2.44	0.0001*	6.16±1.38	6.57±1.36	0.7190
<b>RBC(x10<sup>12</sup>/L)</b>	Mean±SD	4.90±0.72	4.28±0.31	0.0001*	4.90±0.72	4.56±0.71	0.0780
<b>HB (g/dl)</b>	Mean±SD	12.06±1.67	10.36±1.77	0.0001*	12.06±1.67	11.14±1.85	0.0780
<b>PCV (%)</b>	Mean±SD	36.75±5.41	31.06±5.10	0.0001*	36.75±5.41	33.72± 6.20	0.0550
<b>ESR (mm/h)</b>	Mean±SD	6.27 ± 1.09	54.36±39.01	0.0001*	6.27 ± 1.09	61.44±36.76	0.0001*
<b>MCV (fl)</b>	Mean±SD	74.90±5.80	72.84± 8.20	0.4900	74.90±5.80	74.28± 5.32	0.0001*
<b>MCH (pg)</b>	Mean±SD	24.71±1.93	24.02± 2.81	0.5570	24.71±1.93	24.39± 1.72	0.9350
<b>MCHC (g/dl)</b>	Mean±SD	329.24±6.95	330.53±7.87	0.8820	329.24±6.95	328.39±6.96	0.963

**Key:** WBC: White Blood Count, RBC: Red Blood Count, HB: Hemoglobin concentration, PCV: Packed Cell Volume, ESR: Erythrocyte Sedimentation Rate, MCV: Mean Cell Volume, MCH: Mean Cell Hemoglobin, MCHC: Mean Cell Hemoglobin Concentration. \*Significant value

**Table 1c. Post Hoc analysis of Control/ second line and New cases / First line subjects**

Parameter		Control Subjects	Second line	p-value	New cases	First line	p-value
	N =	58	66		38	36	
<b>WBC(x10<sup>9</sup>/L)</b>	Mean±SD	6.16±1.38	5.69±1.56	0.4920	8.67±2.44	6.57±1.36	0.000
<b>RBC(x10<sup>12</sup>/L)</b>	Mean±SD	4.90±0.72	4.60±0.57	0.0690	4.28±0.31	4.56±0.71	0.2870
<b>HB (g/dl)</b>	Mean±SD	12.06±1.67	11.44±1.40	0.2390	10.36±1.77	11.14±1.85	0.2400
<b>PCV (%)</b>	Mean±SD	36.75±5.41	34.97±1.18	0.2950	31.06±5.10	33.72± 6.20	0.1770
<b>ESR (mm/h)</b>	Mean±SD	6.27 ± 1.09	58.03±36.53	0.0001*	54.36±39.01	61.44±36.76	0.8150
<b>MCV (fl)</b>	Mean±SD	74.90±5.80	78.42±5.52	0.0240*	72.84± 8.20	74.28± 5.32	0.0001*
<b>MCH (pg)</b>	Mean±SD	24.71±1.93	24.43±2.44	0.9010	24.02± 2.81	24.39± 1.72	0.9210
<b>MCHC (g/dl)</b>	Mean±SD	329.24±6.95	321.03±8.18	0.0001*	330.53±7.87	328.39±6.96	0.6880

**Key:** WBC: White Blood Count, RBC: Red Blood Count, HB: Hemoglobin concentration, PCV: Packed Cell Volume, ESR: Erythrocyte Sedimentation Rate, MCV: Mean Cell Volume, MCH: Mean Cell Hemoglobin, MCHC: Mean Cell Hemoglobin Concentration. \*Significant value

**Table 1d. Post Hoc analysis of New cases / Second line subjects and First line/Second Line**

Parameter		New cases	Second line	p-value	First line	Second line	p-value
	N =	38	66		36	66	
<b>WBC(x10<sup>9</sup>/L)</b>	Mean±SD	8.67±2.44	5.69±1.56	0.0001*	6.57±1.36	5.69±1.56	0.0960
<b>RBC(x10<sup>12</sup>/L)</b>	Mean±SD	4.28±0.31	4.60±0.57	0.0870	4.56±0.71	4.60±0.57	0.9890
<b>HB (g/dl)</b>	Mean±SD	10.36±1.77	11.44±1.40	0.0150*	11.14±1.85	11.44±1.40	0.8470
<b>PCV (%)</b>	Mean±SD	31.06±5.10	34.97±1.18	0.0040*	33.72± 6.20	34.97±1.18	0.7130
<b>ESR (mm/h)</b>	Mean±SD	54.36±39.01	58.03±36.53	0.9540	61.44±36.76	58.03±36.53	0.9640
<b>MCV (fl)</b>	Mean±SD	72.84± 8.20	78.42±5.52	0.0010*	74.28± 5.32	78.42±5.52	0.0001*
<b>MCH (pg)</b>	Mean±SD	24.02± 2.81	24.43±2.44	0.8850	24.39± 1.72	24.43±2.44	1.0000
<b>MCHC (g/dl)</b>	Mean±SD	330.53±7.87	321.03±8.18	0.0001*	328.39±6.96	321.03±8.18	0.0001*

**Key:** WBC: White Blood Count, RBC: Red Blood Count, HB: Hemoglobin concentration, PCV: Packed Cell Volume, ESR: Erythrocyte Sedimentation Rate, MCV: Mean Cell Volume, MCH: Mean Cell Hemoglobin, MCHC: Mean Cell Hemoglobin Concentration. \*Significant value

Table 1d. is the Post-hoc analysis of New cases / Second line and First line/Second Line subjects. In the first part of the table, the WBC had a significant p value of p= 0.0001 when their Mean±SD was compared (8.67±2.44/5.69±1.56). Other parameters with significant p values are HB p= 0.0150, PCV p= 0.0040, MCV, p= 0.0001 and MCHC, p=0.0001. The rest which are RBC, ESR and MCH all had insignificant p values

of p= 0.087, 0.9540 and 0.8850 respectively. On the second part of the table (First line/Second), the post-hoc reveals that only two out of the eight parameters had significant values which are p= 0.0001 for MCV and p= 0.0001 for MCHC. Other post hoc values are p= 0.0960 for WBC, p= 98900 for RBC, p= 0.8470 for HB, p= 0.7130 for PCV and p=1.0000 for MCH

## DISCUSSION

TB disease is a serious disease of public health concern in third world countries. Much work has not been done about the effects of anti-TB drugs on the hematological parameters in female pulmonary TB patients undergoing DOTS therapy in Nigeria and more so, in the south-south region of the country, Bayelsa State to be precise. The most effective treatment for tuberculosis is the multidrug approach involving a combination of Isoniazid (INH), Rifampin (RIF), Pyrazinamide (PZA) and Ethambutol (EMB) to hamper drug resistance. This blend is highly efficient in treating tuberculosis, but may also lead to toxicity and side effects.

In this research work, a longitudinal approach was taken to check the effects of anti-TB therapy. Control, non-TB infected subjects, newly diagnosed TB subjects, those at the end of First line (intensive phase) treatment and those at the end of second line treatment (Continuation Phase) were all considered. The total white blood cell count (WBC) increased from the control to the newly infected but steadily declined during the treatment period. The ANOVA result gave a significant p value of  $p=0.0001$ . Also, the decline in the number of WBCs from the new cases to the second line treatment revealed a significant post- hoc p value between new cases/ first line ( $p=0.0001$ ) and new case/second line ( $p=0.0001$ ) The post-hoc analysis gave an insignificant p value ( $p=0.4920$ ) between control group and end of treatment group (second line). There was a reduction in number towards normal as treatment continues. The increase WBC in the newly infected subjects has been attributed to an immune response to the TB bacilli. This finding correlates with that of Singh *et al*, 2001,(21). The World Health Organization reports that in tuberculosis disease there is constantly a raised count of white cells beyond normal values

(leucocytosis). Anti-TB drugs such INH and RIF are known to bring about acute leucocytosis either singly or in combination through an unfamiliar mechanism (22) . The increase encountered may be due to a blend of the infection and drugs.

In contrast to WBC, the values for HB, PCV, RBC, MCV and MCH all decreased from control subjects to newly infected subjects and then began to rise steadily as treatment progresses.

The HB, PCV and RBC of the Newly infected pulmonary tuberculosis patients ( $10.36 \pm 1.77$ g/dl,  $31.06 \pm 5.10$  % and  $4.28 \pm 0.31 \times 10^{12}/L$ .) was significantly lower ( $p < 0.05$ ) than that of control

subjects ( $12.06 \pm 1.67$ g/dl,  $36.75 \pm 5.41$ % and  $4.90 \pm 0.72 \times 10^{12}/L$ ). On the other hand, the MCV and MCH lower values were not significant from the control subjects ( $p > 0.05$ ). The value for MCHC for control is  $329.24 \pm 6.95$  and is lower than the Newly infected patients with a value of  $330.53 \pm 7.87$  but not significant ( $p > 0.05$ ). In 2005, Ajayi *et al* (23), in their research work reported significant decreases in these parameters of TB patients. A reduction in haemoglobin concentration and by implication PCV, RBC alongside the absolute values (MC, MCH and MCHC) is known as anemia and have been reported in some studies (24,25). The anemia has been classified as normocytic normochromic with all the features of anemia of chronic disorders (26). Ajayi *et al*. 2005, further states that Pulmonary tuberculosis disease being a chronic disorder, impacts on the hemopoietic system leading to a decrease in erythropoiesis. Various pathogenesis has been advanced in TB-associated anemia, but most investigators have shown suppression of erythropoiesis by inflammatory mediators as a cause of anemia(18) . The drug Rifampin is said to cause erythrocyte death stimulation of  $Ca^{2+}$  permeable cation channels, which

facilitates  $\text{Ca}^{2+}$  entry into RBC from extracellular spaces leading to ceramide production which leads to phagocytosis and apoptosis (27)(28).

It is worthy to note that though there was a decrease in these parameters on the New cases, they were still higher than values from other research works such as that of Ajayi et al 2005 and Nwankwo et al 2005) in Kano and Benin city. Two reasons can be advanced for this; (1) people in this area practice self-medications a lot and the patent medicine stores never hesitates to add hematinics to medicines for every ailment either in the form of capsule or in tonic form. It is believed therefore that these TB patients takes a lot of them as soon as they feel sick. (2) Foods rich in iron are abundant in this area. They include sea foods such as crayfish, fish, snails, periwinkle, prawns etc. which are part of our daily diet. This finding thus denotes that diet of patients to a larger extent plays an important role in the outcome of their hemoglobin concentration.

When RBCs values were compared, a reduction in number was observed on those on 1<sup>st</sup> line drugs when compared with normal (control) values, however there was an increase on the value of the second line regiment ( $4.90 \pm 0.72$ ,  $4.56 \pm 0.71$  and  $4.60 \pm 0.57$  for control, first line and second line respectively). An insignificant p value was obtained in the post-hoc values of the control and first line subjects ( $p = 0.0780$ ). Similarly, the post hoc of the control and second line regiment yielded insignificant result ( $p=0.0690$ ). All other post hoc values were also insignificant; i.e new cases/first line  $P=0.2870$ , new cases /second line  $p=0.0870$  and  $p=0.9890$  for first line/second line. This result is in consonance with a research by Eyuel *et al* 2016, (18). They reported an increase of RBCs and suggests no negative erythropoietic effect of drugs on the bone marrow since the mean value of the first

and second line patients are building up towards normal value meaning recuperation of the subject. The post hoc for HB and PCV followed the same trend with that of RBC save for the post hoc of new cases/second line. They both have significant p values of  $p=0.0150$  and  $0.0040$  for HB and PCV respectively. The similar results of the trio are expected as they are both interconnected. The mean values of ESR rose from control to first line treatment before coming down at the end of the therapy ( $6.27 \pm 1.09$ ,  $54.36 \pm 39.01$ ,  $61.44 \pm 36.76$  and  $58.03 \pm 36.53$  mm/h respectively). The ESR values of new cases ( $54.36 \pm 39.01$  mm/h;) obtained in this study were significantly higher than control values ( $6.27 \pm 1.09$  mm/h),  $p=0.0000$ . This agrees with previous findings (13, 23, 24, 29). The test is usually carried out as a nonspecific test for a range of pathological conditions which include: systemic inflammatory conditions, acute or chronic infections and neoplastic conditions. In all these illnesses, ESR is mostly elevated. Increase in ESR could be attributed to increased production of acute phase proteins and release of proteins by the causative organism (*M.tuberculosis*) into the circulation (16). Comparison of mean values of ESR with ANOVA gave a significant result of  $p=0.0000$ . Similarly, the post hoc gave a  $p=0.0000$  for control/first line and  $p=0.0000$  for control/second line treatment group. However, all the other post-hoc gave insignificant p values. In this study, the cellular components of blood (which contributes partly to the rate of fall) were seen to be getting close to normal values at the end of treatment. Though the value of ESR is also coming down towards normal at the end of the continuation phase, the significant p value ( $p=0.0000$ ) in this line of treatment when compared to the control value could be ascribed to other factors such as the constituents of plasma components which were not considered in this study.

This present study shows a reduction in values of MCV and MCH from control to new cases, and then an increase to the second line regimen. The same was not the case for MCHC as the mean value increased from the control to the new cases and then a decrease through the first line to the second line regimen. Despite this no significant differences ( $p>0.05$ ) were gotten from the post hoc between control and other parameters except for MCV and MCHC in the second line of treatment. The values of the two treatment groups (first line/second line) when compared was significant for MCV and MCHC. Shareef, 2012 and Shafee, 2016, (15,30) also had similar findings in their works on tuberculosis infection. Other research reports reveals that after anti-tuberculosis therapy (DOTS) with these three combination of drugs, streptomycin, rifampicin and isoniazid, the red cell indices MCV, MCH and MCHC were positively affected and reached close to normal values. RBC morphology in pulmonary TB patients is also seen to be generally normocytic normochromic type and during medication, the blood film showed normochromic pictures (31,32,33). This suggests the positive effects of tuberculosis therapy on the restoration of RBC morphology.

#### **CONCLUSION:**

The hematological parameters under study in female Tuberculosis patients at the end of their treatment (DOTS chemotherapy) were not significantly different from those of the control group (i.e. the value of the control compared with that of the second line treatment) except for three parameters (ESR, MCV and MCHC). While our study showed that TB disease significantly affected all the parameters, there was a real improvement in the treatment stage as all parameters were returning back to normal values. This suggests that as time progresses total normalization can occur in all parameters. It

is inferred from this that the DOTS chemotherapy has no negative effect on the hematopoietic process. We therefore suggest further studies that will look into the plasma components of patients undergoing DOTS chemotherapy.

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#### **REFERENCES**

1. Ochei J, Kolhatkar A. Medical Laboratory Science: Theory and Practice. Tata McGraw-Hill publishing company limited, New Delhi.; 2007. 473–479 p.
2. Cobalt EI and Waker N. The growing burden of tuberculosis: a global trend and interactions with the HIV epidemic. Arch Intern Med. 2013;163:1009–21.
3. Blumberg HM, Watkins DL, Berscling JD, Antle A, Moore P and White N. Preventing the nosocomial transmission of tuberculosis. Ann Intern Med. 1995;122:658–63.
4. Ravilion MC and Pio A. Evolution of WHO policies for tuberculosis control, 1948-2001. Lancet; 2002;359:775–80.
5. WHO. Global tuberculosis control: a short update to the 2009 report [Internet]. 2009 [cited 2019 Aug 23]. Available from: [http://www.who.int/tb/publications/global\\_report/2009/en/index.html](http://www.who.int/tb/publications/global_report/2009/en/index.html)
6. Murray C, Styblo K, Rovilion A. Tuberculosis in developing countries,

- burden intervention and cost. . Int J Tuberc Lung Dis. 1990;65:6–24.
7. WHO. The stop TB strategy, Building on and enhancing DOTS to meet the TB-related millennium development goals. 2006.
  8. Kim JY, Shakow A, Casto A, Vande C, and Farmer P. Tuberculosis Control. In: WHO: Trade, Foreign Policy, Diplomacy, and Health. [Internet]. 2017 [cited 2020 Jul 5]. Available from: [http://www.who.int/trade/distance\\_learning/gpgh/gpgh3/en/index9.html](http://www.who.int/trade/distance_learning/gpgh/gpgh3/en/index9.html)
  9. Nigeria. FM of H. NTBLCP Annual Report . 2014 [cited 2018 Aug 20]. Available from: [http://www.health.gov.ng/doc/NTBLCP\\_2014\\_Annual\\_report-2.pdf](http://www.health.gov.ng/doc/NTBLCP_2014_Annual_report-2.pdf)
  10. Ramakrishnan K, Shenbagarathai A R. Kavitha, K, Thirumalaikolundusubramanian P. Hematological Parameters in Pulmonary Tuberculosis Patients with and without HIV infection. Int J Biol Med Res. 2016;7(3):5640–3.
  11. Rohini K, Bhat MS, Srikumar PS, Kumar AM. Assessment of Hematological Parameters in Pulmonary Tuberculosis Patients. Indian J Clin Biochem. 2016;31(3):5640–3.
  12. Sumaira I, Umbreen A, Muhammad AK. Hematological Parameters Altered in Tuberculosis. Pakistan J Physiol. 2015;11(1):13–6.
  13. Atiegha C, Suama P, Daw MOE, Ernest M, Allison M, Obele R, & Mercy I. Some Haematological Indices of Tuberculosis Patients attending Tuberculosis and Leprosy Hospital Igbogene, Bayelsa State, South-South Nigeria. Niger J Heal Allied Res. 2016;3(1):1–8.
  14. Atomsa D, Abebe G, Sewunet T. Immunological Markers and Hematological Parameters among Newly Diagnosed Tuberculosis Patients at Jimma University Specialized Hospital. Ethiop J Heal Sci. 2014;24(4):311–8.
  15. Shareef H. Abnormalities of hematological parameters in newly diagnosed Pulmonary tuberculosis patients in Kirkuk city. Pakistan J Med Sci. 2012;20(5):1486–92.
  16. Akpan PA, Akpotuzor JO, Akwiwu EC. Some Hematological Parameters of Tuberculosis (TB) Infected Africans: the Nigerian Perspective. J Naural Sci Res. 2012;2(1):50–6.
  17. Kutiyal AS, Gupta N, Garg S, Hira HS. A study of haematological and haemostasis parameters and hypercoagulable state in tuberculosis patients in Northern India and the outcome with anti tubercular therapy. J Clin Diagnostic Res. 2017 Feb 1;11(2):OC09-OC13.
  18. Eyuel K, Bamlaku E, Aschalew G, Baye G. Effect of anti-tuberculosis drugs on hematological profiles of tuberculosis patients attending at University of Gondar Hospital, Northwest Ethiopia. Biomedcentral Hematol. 2016;16(1):1–11.
  19. Diatron. Abacus 380 3-Part WBC Differential Analyzer [Internet]. 2017 [cited 2020 Feb 13]. Available from: <http://www.diatron.com/hematology-analyzersabacus-380-3-part-differential-hematology-analyzer>
  20. Inc BC. Coulter HmX Hematology Analyzer and Hmx Hematology Analyzer with Autoloader Documetation. Kraemer Blvd. Brea, California.: Beckman Coulter, Inc., 250 S.; 2011.
  21. Singh KJ, Ahluwalia G, Shamar SK, Saxena R, Chaudhary VP. AM. Significance of haematological manifestations in patients with tuberculosis. Journ Asso phys.

- 2001;49:788–94.
22. Tousif S, Ahmad S, Bhalla K, Moodley P, Das G. Challenges of Tuberculosis Treatment with DOTS: An Immune Impairment Perspective. *J Cell Sci Theory*. 2015;6(23).
23. Ajayi O, Famodu A, Onyemairo J, Iyere C, Onaghise V, Adogun C. Haemorrhological alterations in Nigerian pulmonary tuberculosis patients. *15th European Congress of Clinical Microbiology and Infectious Diseases Copenhagen, Denmark 2005*.
24. Robson SC, White NW, Aronson I, Woollgar R, Goodman H, Jacobs P. Acute phase response and the hypercoagulable state in pulmonary tuberculosis. *British Journal of Hematology*, 1996;93, 943–949.
25. Turken O, Kunter E, Sezer M, Solmazgul E, Cerrahoglu K, Bozkanat E, Ozturk A. & Ilvan A. Hemostatic changes in active pulmonary tuberculosis. *International Journal of Tuberculosis and Lung Diseases*, 2002; 6(10), 927–932.
26. Nwankwo EOK, Kwaru A, Ofulu & Babashani, M.. Haematological changes in tuberculosis in Kano, Nigeria. *Journal of Medical Laboratory Science*, 2005;14, 2, 35-39.
27. Lang F, Gulbins E, Lang P, Zappulla D, Föllner M. Ceramide in suicidal death of erythrocytes. *Cell Physiol Biochem*. 2010;26(1):21–8.
28. Ukpe IS, Southern I. Erythrocyte sedimentation rate in active tuberculosis with or without HIV-coinfection. *South African Med J*. 2006;96(5).
29. Ibeneme EO, Asuquo AE. & Abia-Bassey LN. Prevalence of pulmonary tuberculosis and HIV co-infection among prisoners in Calabar, Nigeria. *Mary Slessor Journal of Medicine*, 2009; 9,2, 10-18.
30. Shafee M, Abbas F, Ashraf M, Mengal MA, Kakar N, Ahmad Z, Ali F. Hematological Profile and Risk Factors Associated with Pulmonary Tuberculosis Patients in Quetta, Pakistan. *Pakistan J Med Sci*. 2014;30(1):36–40.
31. Baynes R, Flax H, Bothwell T, Bezwoda W, MacPhail A, Atkinson P et al. Haematological and iron related measurements in active pulmonary tuberculosis. *Scandinavian J Haematol*. 1986;36(3):280–7.
32. Lombard E & ME. Haematological changes associated with military tuberculosis of the bone marrow. *Tuberc Lung Dis*. 1993;74(2):131–5.
33. E D. Pattern of some haematological indices in newly diagnosed pulmonary tuberculosis cases in Iwo, Nigeria: diagnostic and therapeutic implications. *Niger J Med*. 2000;10(1):18–20.